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**(54) Specific binding assays and reagents therefore**

(57) A sensitive assay method has been discovered that reduces the amount of non-specific binding present in an assay, the method comprising detecting an analyte present in a sample through a specific binding reaction in which either an analogue of the analyte or a specific binding partner of the analyte is immobilized on a solid-phase and said specific binding reaction produces a detectable product immobilized on said solid-phase that

may be correlated to the amount of analyte present in the sample wherein said assay employs an effective amount of a surfactant selected from the group consisting of a polyoxyethylene-alkylether, a polyalkylene oxide-modified polydimethylsiloxane block copolymer, a polyalkylene oxide-modified polymethylsiloxane block copolymer, and mixtures thereof to reduce non-specific binding.

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## Description

This invention relates to heterogeneous specific binding assays.

Biochemical binding assays have been widely utilized in recent years to determine the presence and the concentration of analytes in biological specimens. As known in the art, heterogeneous assays are those in which one of the reactive binding partners is bound to a solid-phase. The various types of heterogeneous assays include, for example, the sandwich method, the indirect method, and the competitive method.

As used herein, "binding partner" is defined to include a specific binding partner to the analyte, an analyte analogue, and the analyte. "Non-specific binding" is defined as non-specific interactions of the binding partners with a solid surface in an assay system.

The non-specific binding often reduces the sensitivity of the heterogeneous assays. The sensitivity of the assay typically refers to the smallest mass of analyte that generates a statistically significant change in the signal generated by the assay when compared to the signal reading obtained in the absence of the analyte. There is a need to develop methods to increase the sensitivity of binding assays (i.e. detect smaller amounts of analyte). Furthermore, a high sensitivity assay is typically capable of an overall higher precision measurement of the analytes.

A number of methods are known for reducing the non-specific binding in heterogeneous assays. For example, proteins such as, bovine serum albumin (BSA), gelatin, and casein, have been added to the assay reagents or preadsorbed on the solid-phase in order to block non-specific adsorption sites. Additionally, the use of various surfactants, often in high concentrations, has been reported in the literature.

While known techniques may assist in reducing some non-specific adsorption, many of the prior art techniques have been associated with interference of the desired specific interaction of the binding partners or lead to the displacement of the specific binding complex formed. Additionally, despite the use of high concentrations of protein and surfactant taught by the literature, a considerable amount of non-specific binding typically still exists in many heterogeneous assays. Alternative means to reducing non-specific binding in heterogeneous assays are needed.

The problem noted above has been solved with the discovery of a method for detecting or measuring an analyte in a sample, the method comprising (1) reacting a solid phase having attached thereto the analyte, an analogue of the analyte or a specific binding partner with a labeled analyte, labeled analyte analogue or labeled binding partner through a specific binding reaction to produce a detectable product immobilized on said solid-phase; and (2) correlating the amount of detectable product with the amount of analyte present in said sample, wherein a surfactant selected from the group consisting of a polyoxyethylene-alkylether, a polyalkylene oxide-modified polydimethylsiloxane block copolymer, a polyalkylene oxide-modified polymethylsiloxane block copolymer, or a mixture thereof, is present in an effective amount to reduce non-specific binding at least about several fold as compared with assays not employing said surfactants in said effective amount.

It has been discovered that the presence of relatively small amounts of the above-identified surfactants in one or more reagents of the assay reduces non-specific binding associated with solid-phase binding assay techniques. Further, the invention may be used to reduce non-specific binding of various types of proteins and oligonucleotides to the solid-phase with only minimal interference to the desired specific interaction. The inventive method may be used in sandwich, indirect, direct and competitive assay formats. Also included is a kit for carrying out the invention.

According to the invention, in a specific binding reaction at least one of the binding partners is immobilized on a solid-phase. The solid-phase may be prepared from inorganic and/or organic materials that will maintain their structural integrity when exposed to water or biological fluids and are widely available commercially. Suitable inorganic materials that may be used in preparing the solid-phase include, but are not limited to, siliceous materials, silica, bentonite, wollastonite, cordierite, and nonsiliceous metal oxides (including magnetic substances, such as iron oxides, ferrite, nickel oxides, cobalt oxides, and the like). Suitable organic materials that may be used include, but are not limited to, synthetic polymers (such as, for example, polystyrene and derivatives thereof, acrylic polymers, polymethacrylates, polyolefins, halogen-containing polymers, polyester, polyurethanes, and polyamides), natural polymers (such as, for example, polysaccharides, cellulose, dextran, agarose, polypeptides, and proteins) or paper, and the like. Combinations of the organic materials and inorganic materials described above may also be used in preparing the solid-phase. For example, magnetic particles with polymeric coatings around metal oxide cores (such as those described in U.S. Patent 4,554,088, Whitehead, et al, 1985, hereby incorporated by reference) may be used. The solid-phase may be in a particulate form (varying from a finely divided powder, such as magnetite, to a coarse granular material) or a shaped article (such as a bead, test tube, microtiter plate, cuvette, membrane, film, filter paper, disc, and so on). Although any of the above may be used, more preferably, the solid-phase is an article or particle that comprises a polymeric material (most preferably polystyrene, polypropylene, or mixtures thereof) that is present on at least the surface of said article or particle, including solid-phases made of entirely polymeric material as well as polymer coated (preferably polystyrene) magnetic (preferably iron oxide) particles.

The binding partner immobilized on the solid-phase may be an unlabeled specific binding partner of the analyte of interest, an unlabeled analogue of the analyte of interest, or the analyte of interest. The specific binding partner is

defined as a substance capable of pairing with the targeted analyte or analyte analogue through a specific binding reaction. The analyte analogue is defined as a substance that is capable of pairing with the specific binding partner of the targeted analyte through a specific binding reaction. The specific binding reaction product complex refers either to a complex of the targeted analyte-specific binding partner, a complex of the analyte analogue-specific binding partner or a complex of a first binding partner-analyte-second binding partner "sandwich" (where the first and second binding partners may be the same or different). The pairing of the specific binding partner and the targeted analyte (and analyte analogue, if used) may be through any number of reactions, including immunological, chemical, and complementary binding. The binding partner that is immobilized on the solid-phase is referred to herein as the "solid-phase reagent". The specific binding partner that used in the invention may be, for example, an antibody, an antigen, avidin, biotin, thyroxin, a thyroxin-binding globulin, a polysaccharide, a oligonucleotide, a polynucleotide, phosphorylcholine, aminoethylidihydrogen phosphate residues, estrogen, vitamin B-12 intrinsic factor, lectins, binding proteins, and various other proteins, receptors, and peptides, nucleic acids, nucleosides, and mixtures thereof.

The invention may be utilized in detecting and/or measuring a number of analytes, including proteins, peptides, polyamino acids, oligonucleotides, RNA, DNA polysaccharides and so on, through binding assays, immunoassays, and nucleic acid hybridization assays (i.e. gene probe assays). This method is particularly useful in reducing non-specific binding of binding partners in assays that measure and or detect the analytes described above.

Any number of known means may be used to directly detect or indirectly detect the immobilized specific binding complex in the assay. Preferably the assay is carried out with the use of a "tracer reagent", as commonly used in immunoassays and gene probe assays. As used herein, the "tracer reagent" is defined as a labeled analyte, a labeled analyte analogue or a labeled specific binding partner. The label attached to the analyte, analyte analogue or specific binding partner is a substance capable of producing a detectable signal. Any number of substances may be used as the label, including, for example, radioisotopes, luminescent, fluorescent or chemiluminescent materials, enzymes, liposomes, as well as metal and non-metal materials. The label can be attached to the specific binding partner, analyte, or analyte analogue directly (e.g. covalent bonding) or indirectly (e.g. biotin/avidin, DNP/anti-DNP) by known techniques. The immobilized complex may be detected, directly (by, for example, measuring the label on the complex) or indirectly (by, for example, measuring the unreacted label) by any number of methods, depending upon what is employed as the label substance. Preferably the label is a chemiluminescent label (most preferably acridinium ester) or an enzymatic label. Particularly preferred chemiluminescent labels are acridinium esters (AE), such as those described in U.S. Patent 4,918,192 (hereby incorporated by reference) and EP-A-617,288 where the measurable signal is a chemiluminescent light emission typically activated by an acid reagent (e.g.  $H_2O_2$  in  $HNO_3$ ) followed by a base reagent (e.g.  $NaOH$ ).

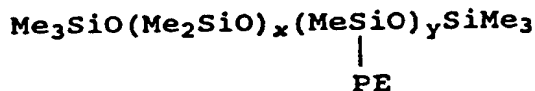
Heterogeneous assays typically require a separation of the solid-phase immobilized specific binding reaction complex from the unreacted tracer reagent. A number of means of separation may be employed, many of which use a liquid solution alone or in combination with other techniques (such as, for example, magnetic separation). When used, the liquid solution is referred herein as a "wash reagent" and is typically an aqueous solution that may contain other constituents known to those skilled in the art, as discussed in more detail hereinafter.

Of the three general classes of surfactants described in the invention, the first class are polyoxyethylene-alkylether nonionic surfactants preferably having the following general structure:



wherein  $n$  is an integer having a value of from about 1 to about 18 and  $m$  is an integer having a value of from about 2 to about 100. These compounds are widely available commercially. Particularly preferred is polyoxyethylene (4) lauryl ether (BRIJ® 30 surfactant, ICI Americas, Delaware).

The second and third classes of surfactants that may be utilized are polyalkylene oxide-modified polyalkylsiloxane block nonionic copolymers (also commonly referred to as silicone glycol copolymers) of two structural types, identified herein as Formulas A and B. The structural type identified by Formula A are linear polydimethylsiloxanes to which polyethers have been grafted through a hydrosilation reaction. This process results in an alkyl-pendant copolymer in which the polyalkylene oxide groups are attached along the siloxane backbone through a series of hydrolytically stable Si-C bonds. These products are shown by Formula A as follows:



A

wherein PE =  $-CH_2CH_2CH_2O(EO)_p(PO)_qZ$ , Si represents silicon, Me represents methyl, EO represents ethyleneoxy, PO represents 1,2-propyleneoxy, and Z can be either hydrogen or a lower alkyl  $C_1$ - $C_4$  radical,  $o$  is an integer from about 4 to 600, and  $p$  is an integer from about 4 to 100, and  $x$  and  $y$  are integers having a value of from about 1 to about 10. The polyalkylene oxide-modified polydimethylsiloxane copolymers represented by Formula B, below, are branched polydimethylsiloxanes to which polyethers have been attached through condensation chemistry. This process results in an alkoxy-endblocked copolymer in which the polyalkylene oxide groups are attached at the ends of the

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silicone backbone through Si-O-C bonds. These products are shown by Formula B as follows:



wherein PE =  $-(\text{EO})_s(\text{PO})_t\text{R}$ , R is a  $\text{C}_1\text{-C}_4$  lower alkyl group,  $r$  and  $q$  are integers from about 3 to about 50,  $s$  is an integer from about 5 to 100,  $t$  is an integer from about 5 to about 100, and Me, Si, EO, and PO are as defined previously. Both classes of these block polymers are not usually present as single chemical compounds but rather are a mixture of several individual compounds each having a different molecular weight and a different molar ratio of polyethylene oxide groups (EO) to polypropylene oxide groups (PO), thus the values for  $o$  and  $p$  and  $s$  and  $t$  are average values which can be determined for the mixture from the molecular weight and the molar ratio of EO/PO. Additionally the values of  $x$  and  $y$  and  $r$  and  $q$  are also average values that may be determined by the molecular weight of the copolymers. Both classes of block copolymers are widely commercially available such as those manufactured by OSi Specialties under the SILWET® L surfactants series, with SILWET® L-7604 surfactant (polyalkylene oxide modified polydimethyl siloxane) and SILWET® L-7607 surfactant (polyalkylene oxide modified polymethyl siloxane) most preferred.

The usual constituents known to those skilled in the art may be included in the reagent mixtures (i.e. tracer, solid-phase, and wash reagents). Constituents may include buffer substances such as phosphate buffer, citrate buffer, borate buffer and/or bovine serum albumin and/or preservatives and the like. Particularly preferred in tracer reagents and wash reagents is a phosphate buffer and bovine serum albumin.

The identified surfactants may be utilized in different (or all) of the defined reagent mixtures (i.e. solid-phase, tracer, and/or wash reagents) to reduce non-specific binding associated with heterogeneous assays. Any number of combinations may be practiced. When employed in the solid-phase reagent, the surfactant may be included in a solution used to pretreat the solid-phase prior to the attachment of the binding partner to the solid-phase and/or included in the solid-phase reagent after the binding partner is attached to the solid-phase.

Surprisingly, the selected surfactants reduce non-specific binding with minimal to no interruption of the specific binding reaction and specific binding complex on the solid-phase, even when the surfactant is present in the wash reagent. An "effective amount" of the surfactant(s) is defined as an amount of about 0.001 weight/volume percent to about 3 weight/volume percent, with weight/volume (w/v) percentages based on the total volume of each reagent mixture that includes the selected surfactants. This invention is not limited to utilizing the above-stated amount of surfactant(s). A larger amount of the surfactant may be employed, however, one of the advantages of the invention is that the selected surfactants have been found to be effective in reducing non-specific binding when present in relatively small amounts. More preferably the surfactant is employed in a range from 0.005 w/v percent (most preferably 0.01 w/v percent) to 1.5 w/v percent (most preferably 1 w/v percent). The presence of the surfactants in one or more of the solid-phase, tracer, and/or wash reagent(s) in an effective amount may reduce non-specific binding interactions of the binding partners to the solid-phase in an amount of at least about several fold up (in some instances up to 10,000 fold and more) as compared to the non-specific binding of assays not employing said surfactants. In particularly preferred assays, a reduction of non-specific binding of at least about 1000 fold may be accomplished by including at least one of the defined surfactants to one or more of the tracer, solid-phase, or wash reagent(s).

The invention also includes test kits for immunoassays and gene probe assays comprising (a) a solid-phase reagent; (b) a tracer reagent; and (c) a wash reagent, wherein at least one of said reagents includes a surfactant selected from the group consisting of a polyoxyethylene-alkylether, a polyalkylene oxide-modified polymethylsiloxane block copolymer, a polyalkylene oxide-modified polydimethylsiloxane block copolymer, and mixtures thereof, wherein said surfactant is present in an amount ranging from about 0.001 w/v % to about 3 w/v %. Optionally, the test kits may include calibrator solutions which contain a known amount of analyte, as are well known. When the surfactant is employed in the solid-phase reagent, the surfactant may be used to pretreat the solid-phase prior to the attachment of the binding partner to the solid-phase and/or included in the actual reagent mixture after the binding partner has been attached to the solid-phase.

The invention may be practiced in a number of assay formats, including those described below.

### Labeled analyte or labeled analyte analogue, competitive format:

A heterogeneous specific binding assay for detecting or measuring an analyte in a sample, the assay comprising (1) contacting the sample with (a) a tracer reagent comprising a labeled analyte analogue or labeled analyte; and (b) a solid-phase reagent comprising a solid-phase having attached thereto a specific binding partner to said analyte or analyte analogue, to form an immobilized specific binding reaction product complex comprising said labeled analyte analogue or labeled analyte paired with said immobilized specific binding partner; (2) separating said solid-phase having the immobilized complex thereon from said tracer reagent by a method comprising employing a wash reagent; (3) directly or indirectly detecting said label on said immobilized complex; and (4) correlating said detected label with the presence of said analyte in said sample, wherein said method includes the presence of a surfactant selected from the group consisting of a polyoxyethylene-alkylether, a polyalkylene oxide-modified polydimethylsiloxane block copol-

mer, a polyalkylene oxide-modified polymethylsiloxane block copolymer, or a mixture thereof in one or more reagents selected from the group consisting of said wash reagent, tracer reagent, or solid-phase reagent, said surfactant present in an effective amount to reduce non-specific binding in said assay at least about several fold.

#### Labeled specific binding partner, competitive format:

A heterogeneous specific binding assay for detecting or measuring an analyte in a sample, the assay comprising (1) contacting the sample with (a) a tracer reagent comprising a labeled specific binding partner of the analyte; and (b) a solid-phase reagent comprising a solid-phase having attached thereto an analyte or an analogue analyte, to form an immobilized specific binding reaction product complex comprising said labeled specific binding partner paired with said immobilized analyte or analogue analyte; (2) separating said solid-phase having the immobilized complex thereon from said tracer reagent by a method comprising employing a wash reagent; (3) directly or indirectly detecting said label on said immobilized complex; and (4) correlating said detected label with the presence of said analyte in said sample, wherein said method includes the presence of a surfactant selected from the group consisting of a polyoxyethylene-alkylether, a polyalkylene oxide-modified polydimethylsiloxane block copolymer, a polyalkylene oxide-modified polymethylsiloxane block copolymer, or a mixture thereof in one or more reagents selected from the group consisting of said wash reagent, tracer reagent, or solid-phase reagent, said surfactant present in an effective amount to reduce non-specific binding in said assay at least about several fold.

#### Sandwich assay, noncompetitive format:

A specific binding sandwich assay for detecting or measuring an analyte in a sample, the assay comprising (1) contacting the sample with (a) a tracer reagent comprising a labeled specific binding partner of the analyte; and (b) a solid-phase reagent comprising a solid-phase having attached thereto an unlabeled specific binding partner to the analyte to form an immobilized specific binding reaction product complex comprising said labeled specific binding partner paired with said analyte paired with said immobilized unlabeled specific binding partner; (2) separating said solid-phase having the immobilized complex thereon from said tracer reagent by a method comprising employing a wash reagent; (3) directly or indirectly detecting said label on said immobilized complex; and (4) correlating said signal with the presence of said analyte in said sample, wherein said method includes the presence of a surfactant selected from the group consisting of a polyoxyethylene-alkylether, a polyalkylene oxide-modified polydimethylsiloxane block copolymer, a polyalkylene oxide-modified polymethylsiloxane block copolymer, or a mixture thereof in one or more reagents selected from the group consisting of said wash reagent, tracer reagent, or solid-phase reagent, said surfactant present in an effective amount to reduce non-specific binding in said assay at least about several fold.

In one preferred aspect of this invention, the assay is carried out utilizing polyalkylene oxide modified polydimethylsiloxane in the tracer reagent and polyoxyethylene lauryl ether or polyalkylene oxide modified polymethyl siloxane in the wash reagent.

Alternatively, the above-described surfactants may be used in assay systems that employ a solid phase, but do not employ a separation step (termed pseudo-homogeneous assays herein). In pseudo-homogeneous assays a method of reducing non-specific binding is provided, the method comprising employing one or more of the above-described surfactant(s) in an effective amount in the assay to reduce non-specific binding at least about several fold.

Preferably, the surfactants are present in a solution used in such assays, for example the solution wherein the sample is introduced, or (if used) in the tracer reagent, and/or solid phase reagent (if used as a separate reagent).

#### EXAMPLES

Materials that were used in the Examples included BRIJ® 30 surfactant [polyoxyethylene (4) lauryl ether]; TWEEN® 20 surfactant (polyoxyethylene sorbitan monolaurate), polyethylene oxide (PEO, Cat. No. P 6667, average MW 10,000) and PVA (polyvinyl alcohol, Cat. No. P 8136, average MW 30,000-70,000), bovine gamma globulin (BgG), and hemoglobin (HB), all of which were all purchased from Sigma Chemical Co. (St. Louis, MO). SILWET® L-7604 (polyalkylene oxide modified polydimethyl siloxane) and SILWET L-7607 (polyalkylene oxide modified polymethyl siloxane) were obtained from OSi Specialties, Inc. TRITON® X-100 and TRITON X-705 surfactants (octyl phenol polyether alcohols) were obtained from Union Carbide Corp. PLURONIC® poly(oxyethylene-oxypropylene) copolymer surfactants and TETRONIC® alkoxylated diamine surfactants were obtained from BASF Corp. (Parsippany, NJ). Polystyrene tubes, 12x75 mm, were purchased from Sarstedt Inc. (Cat. No. 55.476, Pennsauken, NJ). Polypropylene cuvettes, 12x75 mm, were obtained from Becton Dickinson Corp. (Falcon 2002, Lincoln Park, NJ). Polypropylene cuvettes were manufactured for Ciba-Corning Diagnostics Corp. (Medfield, MA). Polystyrene-encapsulated magnetic particles were purchased from Bangs Laboratories, Inc. (Stock No. M0009400CN, Carmel, IN). The monoclonal antibodies against triiodothyronine, creatine kinase, dinitrophenol, and thyroid stimulating hormone were mouse antibodies prepared with

techniques well-known to those skilled in the art. The acridinium ester (AE) label was conjugated to the various specific binding partners (thus forming the tracer) in a manner as described in I. Weeks, et al., "Acridinium Esters as High-Specific Activity Labels in Immunoassay", *Clin. Chem.*, 29, 1474-1479 (1983). All weight/volume percentages of the surfactants are based on the total volume of each reagent mixture that includes the surfactant. The AE-labeled oligonucleotide was a 5'-NH<sub>2</sub>-mecA-495 where the sequence was 5' GAA GAT GGT ATG TGG AAG TTA GAT ATT, where A = adenine, G = guanine, and T = thymine.

A magnetic separation rack (available from Ciba Corning Diagnostics Corp., Medfield, Mass.) was used in the separation step. The chemiluminescent labels were activated by first contacting the separated solid-phase with 0.3 ml of Flash Reagent 1 (0.1 N HNO<sub>3</sub> in a 0.5% aqueous solution of H<sub>2</sub>O<sub>2</sub>) followed by contacting the solid-phase with 0.3 ml of Flash Reagent 2 [0.25 N NaOH in a 0.5% aqueous solution of ARQUAD® 16-50 N-alkyl trimethyl ammonium chloride (50% active purchased from AKZO Chemical Inc., Chicago, IL)].

#### EXAMPLE 1

##### Effect of BRIJ 30 and SILWET L-7607 in the Wash Reagent on Non-Specific Binding of Bovine Gamma Globulin (BgG) to Polystyrene Tube

In the first step, the following reagents were added to a 12x75 mm polystyrene tube: (a) 0.1 ml of PBS/BSA buffer [0.05 M sodium phosphate monobasic, 0.15 M sodium chloride, 1 mM ethylenediaminetetraacetic acid (EDTA), 0.02 w/v % sodium azide, 0.1 w/v % bovine serum albumin (BSA), pH 7.4]; (b) 0.1 ml of acridinium ester (AE)-labeled bovine gamma globulin (BgG) at approximately 1x10<sup>8</sup> RLU (relative light unit) in PBS/BSA buffer; and (c) 0.5 ml of PBS/BSA. After each addition, the tube was vortexed.

In the second step, following 1 hour of incubation at room temperature, the content of each tube was decanted and blotted on an absorbent tissue paper for 3 min. Next, a wash solution (1 ml PSA/BSA buffer) was prepared without a surfactant (CONTROL) as well as numerous wash reagents that included 1 ml of PBS/BSA buffer with 0.1 w/v % of the different surfactants shown in TABLE I. Each reagent was added to separate tubes, vortexed, decanted and blotted for 3 min. This wash step was repeated for a total of 4 times. Thereafter, 5 ml of deionized water were added to each tube, decanted, and blotted for 3 min. Lastly, 0.1 ml of deionized water was added to each tube and the non-specific binding of AE-labeled BgG to the polystyrene tube was measured after flashing the AE label by adding 0.3 ml of Flash Reagent 1 followed by 0.3 ml of Flash Reagent 2 whereupon the resulting light emission was measured by a Magic® Lite Analyzer II (Ciba Corning Diagnostics Corp., Medfield, MA). Data measured from each test tube are summarized in TABLE 1. The data shown are presented as fractional non-specific binding defined as Relative Light Units (RLUs) remaining in each tube divided by the input RLU, i.e. 1x10<sup>8</sup>.

TABLE 1

Non-Specific Binding of Acridinium Ester (AE)-Bovine Gamma Globulin (BgG) on Polystyrene Tube		
Surfactant in PSA/BSA Wash Reagent	Amount of Surfactant	Fractional Non-Specific Binding
None (Control)	0	4 x 10 <sup>-4</sup>
BRIJ® 30 <sup>1</sup> . (Invention)	0.1 w/v %	3 x 10 <sup>-8</sup>
SILWET® L-7607 <sup>2</sup> . (Invention)	0.1 w/v %	3 x 10 <sup>-8</sup>
TWEEN® 20 <sup>3</sup> . (Comparative)	0.1 w/v %	2 x 10 <sup>-6</sup>
TRITON® X-100 <sup>4</sup> . (Comparative)	0.1 w/v %	2 x 10 <sup>-7</sup>

1. Polyoxyethylene (4) lauryl ether

2. Polyalkyleneoxide modified polymethyl siloxane

3. Polyoxyethylene sorbitan monolaurate

4. Octyl phenol polyether alcohol

As the results in TABLE 1 show, addition of 0.1 w/v % BRIJ 30 or 0.1 w/v % SILWET L-7607 to the PBS/BSA wash reagent was observed to reduce fractional non-specific binding of AE-BgG to polystyrene tubes more than 10,000 fold. In contrast, addition of 0.1 w/v % of TWEEN 20 or TRITON X-100 to the wash solution was observed to reduce non-specific binding 200 and 2000 fold, respectively.

**EXAMPLE 2****Effect of BRIJ 30 and SILWET L-7607 in the Wash Reagent on Non-Specific Binding of Anti-Thyroid Stimulating Hormone (TSH) Antibody to Polystyrene**

Non-specific binding of a mouse monoclonal antibody against thyroid stimulating hormone (TSH) to polystyrene tube was measured as described in Example 1, with the exception that anti-TSH was substituted for the BgG and the first step utilized a BSA solution of 0.1 ml of 5 w/v % BSA solution (0.05 M sodium phosphate monobasic, 0.02 w/v % sodium azide, 5 w/v % BSA, pH 7.4) instead of the PSA/BSA buffer.

TABLE 2

Non-Specific Binding of AE-Monoclonal Antibody (Anti-TSH) to Polystyrene Tube		
Surfactant in PBS/BSA Wash Reagent	Amount of Surfactant	Fractional Non-Specific Binding
None (Control)	0	$5.5 \times 10^{-4}$
BRIJ® 30 (Invention)	0.1 w/v %	$1 \times 10^{-8}$
SILWET® L-7607 (Invention)	0.1 w/v %	$1 \times 10^{-8}$
TRITON® X-705 (Comparative)	0.1 w/v %	$1.2 \times 10^{-5}$

As demonstrated in TABLE 2, addition of 0.1 w/v % BRIJ 30 or SILWET L-7607 to the wash reagent was observed to reduce fractional non-specific binding of Anti-TSH to polystyrene over 50,000 fold. In contrast, addition of 0.1 w/v % TRITON X-705 to the wash solution (such as taught by U.S. Patent 4,668,639) was observed to reduce the fractional non-specific binding less than 50 fold.

**EXAMPLE 3****Effect of Concentration of BRIJ 30 and SILWET L-7607 in the Wash Reagent on Non-Specific Binding of Various Tracer Reagents**

AE-labeled proteins BgG and hemoglobin (HB) as well as monoclonal antibodies against triiodothyronine, creatine kinase (CK-MB), dinitrophenol, and Thyroid stimulating hormone (TSH) were tracer reagents evaluated for non-specific binding to polystyrene using two different concentrations of various surfactants in the PBS/BSA buffer with the protocol described in Example 2, with the exception that in the second step the tubes were washed three times instead of four. The concentration and types of surfactants tested are shown in TABLES 3 and 4, where the following is a list of abbreviations used: AE = acridinium ester label; BgG = bovine gamma globulin; HB = hemoglobin, Anti-TY 1-3 = three different lines of anti-triiodothyronine antibodies, Anti-CK = anti-creatine kinase antibody, Anti-DNP = anti-dinitrophenol antibody, and Anti-TSH = anti-thyroid stimulating hormone antibody; E = times  $10^x$  (i.e.  $7.7E-8 = 7.7 \times 10^{-8}$ ); Inv. = invention; Comp. = comparison; and SILWET = a polyalkylene oxide modified polymethyl siloxane (SILWET L-7607).

TABLE 3

Effect of BRIJ 30 and Triton X100 on Fractional Non-Specific Binding					
	Wash Reagent	PBS/BSA			
AE-labeled Tracer	Control	BRIJ-30 0.01 w/v% (Inv.)	TRITON X100 0.01 w/v% (Comp.)	BRIJ-30 0.1 w/v% (Inv.)	TRITON X100 0.1 w/v% (Comp.)
BgG	5.4E-5	1.2E-6	5.7E-6	7.7E-8	8.4E-7
HB	6.9E-4	3.1E-6	6.3E-5	1.4E-6	1.1E-6
Anti-TY1	6.5E-6	7.7E-8	2.8E-7	<5E-8	1.1E-7
Anti-TY2	2.1E-5	1.5E-7	3.1E-6	<5E-8	3.1E-7
Anti-TY3	4.2E-5	2.4E-7	2.7E-6	1.1E-7	5.5E-7
Anti-CK	1.4E-5	2.3E-7	3.1E-6	1.9E-7	5.0E-7
Anti-DNP	7.9E-6	3.3E-7	8.3E-7	<5E-8	<5E-8
Anti-TSH	5.5E-4	1.7E-6	1.1E-5	3.4E-7	1.2E-6

TABLE 4

Effect of SILWET L-7607 and TWEEN 20 on Fractional Non-Specific Binding					
	Wash Reagent	PBS/BSA			
AE-labeled Tracer	Control	SILWET 0.01 w/v% (Inv.)	TWEEN-20 0.01 w/v% (Comp.)	SILWET 0.1 w/v% (Inv.)	TWEEN-20 0.1 w/v% (Comp.)
BgG	4.3E-5	2.2E-6	1.8E-6	3.1E-7	7.2E-7
HB	6.2E-4	3.2E-5	2.1E-4	8.2E-7	9.8E-6
Anti-TY1	6.1E-6	2.6E-7	3.1E-7	1.1E-7	4.8E-8
Anti-TY2	2.1E-5	1.8E-6	2.3E-6	<5E-8	1.1E-6
Anti-TY3	3.5E-5	2.6E-6	4.7E-6	3.8E-7	1.5E-6
Anti-CK	1.3E-5	3.8E-6	1.9E-6	3.3E-7	7.7E-7
Anti-DNP	7.3E-6	1.8E-7	4.3E-7	<5E-8	1.4E-7
Anti-TSH	5.4E-4	7.9E-6	1.1E-5	5.8E-7	3.8E-6

As the results summarized in TABLES 3 and 4 illustrate, the average decrease in the non-specific binding of these eight tracers with 0.01 w/v % BRIJ 30 was observed as approximately 150 fold and SILWET L-7607 was approximately 40 fold, compared to 20 and 15 fold reduction obtained with TWEEN 20 and TRITON X-100, respectively. At 0.1 w/v %, the average decrease in non-specific binding for BRIJ 30 was observed as 850 fold. At 0.1 w/v % SILWET L-7607 showed an average decrease in non-specific binding of 602 fold. At 0.1 w/v %, the comparative surfactants, TRITON X-100 and TWEEN 20, showed a 273 and 63 fold, respectively.

#### EXAMPLE 4

##### Effect of Surfactants in Tracer Reagent on Non-Specific Binding of Anti-Creatine Kinase Antibody to Polystyrene

Non-specific binding of an AE-labeled anti-creatine kinase monoclonal antibody to polystyrene tube was measured with the protocol described in Example 2 with several modifications. AE-labeled creatine kinase antibody was tested for the non-specific binding and the surfactants tested (as identified in TABLE 5) were added to the tracer reagent in Step 1 rather than the wash reagent in Step 2. Additionally, the surfactants were employed at a higher concentration, 1 w/v %. Results are summarized in TABLE 5.

TABLE 5

Fractional Non-Specific Binding of AE-Anti-Creatine Kinase Antibody to Polystyrene Tube		
Surfactant in PSA/BSA Tracer Reagent	Amount of Surfactant	Fractional Non-Specific Binding
Control	0	$3.1 \times 10^{-5}$
SILWET® L-7604 <sup>1</sup> (Invention)	1 w/v %	$1.4 \times 10^{-7}$
TETRONIC® 904 <sup>2</sup> (Comparative)	1 w/v %	$2.2 \times 10^{-6}$
TRITON® X-705 <sup>3</sup> (Comparative)	1 w/v %	$8.6 \times 10^{-6}$

1. Polyalkyleneoxide modified polydimethyl siloxane

2. Alkoxyated ethylene diamine

3. Octylphenoxy polyethoxy ethanol

As shown by the data in TABLE 5, the addition of 1 w/v % SILWET L-7604 was observed to reduce non-specific binding of the Anti-Creatine Kinase antibody to polystyrene 200 fold. Addition of 1 w/v % TETRONIC 904 to PBS/BSA resulted in a 10 fold reduction. Addition of 1 w/v % Triton® X-705 was observed to decrease non-specific binding by four fold.

#### EXAMPLE 5

##### Effect of Surfactants in Tracer Reagent on Non-specific Binding of BgG to Polystyrene

Non-specific binding of a AE-labeled BgG to polystyrene tube was measured as described in Example 4 with the modifications that the AE-BgG was substituted in the tracer reagent. Results are summarized in TABLE 6.



TABLE 6

Non-Specific Binding of Acridinium Ester(AE)-BgG Polystyrene Tube		
Surfactant in PSA/BSA Tracer Reagent	Amount of Surfactant	Fractional Non-Specific Binding
Control	0	$3.5 \times 10^{-4}$
SILWET® L-7604 <sup>1</sup> . (Invention)	1 w/v %	$4.6 \times 10^{-6}$
TETRONIC® 904 <sup>2</sup> . (Comparison)	1 w/v %	$2.7 \times 10^{-6}$
PLURONIC® P84 <sup>3</sup> . (Comparison)	1 w/v %	$5.3 \times 10^{-6}$

1. Polyalkyleneoxide modified polydimethyl siloxane  
 2. Alkoxylated ethylene diamine  
 3. Poly(oxyethylene-oxypropylene) copolymer

**EXAMPLE 6**Effect of Surfactants in Wash Reagent on Non-Specific Binding of BgG to Polypropylene

Fractional non-specific binding of AE-BgG to the polypropylene cuvettes was studied as follows: 0.1 ml of 5 w/v % BSA solution described in Example 2 was added to each cuvette. Next, 0.1 ml of AE-BgG at  $1 \times 10^8$  RLU and 0.5 ml of PBS/BSA buffer were added sequentially. After one hour of incubation at room temperature, cuvettes were washed three times with either water ("Control") or a wash reagent comprising PBS buffer (0.05 M sodium phosphate mono-basic, 0.15M sodium chloride, 1 mM EDTA, and 0.02 w/v % sodium azide), 1 w/v % polyvinyl alcohol (PVA), and 0.1 w/v % BRIJ 30 ("PBS/PVA/BRIJ 30") and thereafter flashed with Flash Reagent 1 and Flash Reagent 2 and measured. The RLUs remaining in each cuvette was determined on the ACS:180® instrument (a fully automated, random access, chemiluminescent immunoassay system, as manufactured by Ciba Corning Diagnostics Corp.). The measured RLUs remaining in each cuvette represented the fractional non-specific binding, where the fractional non-specific binding of BgG to polypropylene cuvettes was observed to be reduced from  $1.6 \times 10^{-4}$  to  $7.8 \times 10^{-7}$  when the water wash was replaced with PBS/PVA/BRIJ 30 wash reagent.

**EXAMPLE 7**Effect of BRIJ 30 in the Wash Reagent on Specific and Non-Specific Binding of Creatine Kinase (CK-MB) Assay

Effect of wash reagent composition on specific and non-specific binding of Magic® Lite CK-MB (creatine kinase) assay was studied. 0.1 ml of CK-MB standard, containing either zero or 51.6 ng/ml CK-MB, was added to each polystyrene tube followed by 0.1 ml of AE-labeled anti-CK-MB antibody ( $1.6 \times 10^7$  RLU). Next, 0.5 ml of anti-CK-MB antibody immobilized on paramagnetic iron oxide particles was added. Following one hour of incubation, each tube was washed two times with water; PBS/BSA; or PBS/BSA/ 0.1 w/v % BRIJ 30 and thereafter flashed and measured in the manner described in Example 1. Results are summarized in TABLE 7 below. As shown, addition of BRIJ 30 to the wash reagent improved the signal-to-noise of the assay from 263 to 417.

TABLE 7

Specific and Non-Specific Binding of Creatine Kinase Assay			
Wash Reagent	Creatine Kinase Concentration (ng/ml)		Signal/Noise
	0	51.6	
Water (Control)	1933	302240	156
PBS/BSA (Control)	860	226235	263
PBS/BSA/BRIJ 30 (Invention)	557	232125	417

**EXAMPLE 8**Effect of Surfactant in Wash Reagent on Non-Specific Binding of an Oligonucleotide to Polystyrene

Fractional non-specific binding of AE-labeled oligonucleotide 5'-NH<sub>2</sub>-mecA-495 to polystyrene tubes was significantly reduced by addition of 0.1 w/v % BRIJ 30 to the wash reagent. The protocol of Example 2 was used with three

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washes using either water or PBS/PVA/BRIJ 30. Non-specific binding of the oligonucleotide tracer to polystyrene was reduced from  $1.4 \times 10^{-4}$  to  $< 5 \times 10^{-8}$  when the standard washing solution, water, was replaced with PBS/PVA/BRIJ 30.

### EXAMPLE 9

#### Effect of Surfactant in Wash Reagent on Non-Specific Binding of an Oligonucleotide to Polypropylene

Fractional non-specific binding of AE-labeled oligonucleotide 5'-NH<sub>2</sub>-mecA-495 to polypropylene tubes was reduced from  $5.9 \times 10^{-4}$  to  $1.9 \times 10^{-5}$  when the water wash solution was replaced with a PBS/PVA/BRIJ 30. The modification of the protocol of Example 2 as described in Example 6 was followed.

### EXAMPLE 10

#### Effect of Surfactant in Wash Reagent on Non-Specific Binding of an Oligonucleotide to Polypropylene

The non-specific binding of the AE-labeled oligonucleotide tracer 5'-NH<sub>2</sub>-mecA-495 to polypropylene cuvettes was evaluated using TETRONIC 904 surfactant. The assay protocol was as follows: First, 0.1 ml of 5 w/v % BSA was added to each cuvette followed by 0.1 ml of the 5'-AE-NH<sub>2</sub>-495 in PBS/BSA with or without 0.1 w/v % TETRONIC 904. Next, 0.45 ml of PBS/BSA with or without 0.1 w/v % TETRONIC 904 was introduced into each cuvette. After 7.5 min of incubation at 37°C, the cuvettes were washed 3 times with PBS/0.1 w/v % BRIJ 30. The non-specific binding of the oligonucleotide to the polypropylene cuvettes was reduced from  $1.4 \times 10^{-5}$  to  $5.2 \times 10^{-6}$  when 0.1 w/v % TETRONIC 904 was added to the buffer solutions as flashed and measured in the manner described in Example 6.

### EXAMPLE 11

#### Effect of Surfactants in Wash or Solid-Phase Reagent on Non-Specific Binding of BgG to Polystyrene-Coated Magnetic Particles

In this experiment, the effect of BRIJ 30 and TETRONIC 908 on non-specific binding of AE-BgG to polystyrene encapsulated magnetic particles was studied. First, 0.1 ml of PBS/BSA or PBS containing 1 w/v % TETRONIC 908 was added to a polystyrene tube. Then, 0.5 ml of magnetic particles at 0.1 mg/ml in PBS/BSA was added followed by 0.1 ml of AE-BgG at  $10^8$  RLU. The samples were vortexed following each addition. After one hour incubation at room temperature, samples were washed three times with PBS/BSA or PBS/PVA/BRIJ 30 and thereafter flashed and measured in the manner described in Example 1. Results are summarized in TABLE 8.

TABLE 8

Fractional Non-Specific Binding of AE-Labeled BgG to Magnetic Polystyrene Microspheres		
Wash Reagent	FRACTIONAL NON-SPECIFIC BINDING	
	Solid-phase Reagent:	
	PBS/BSA	PBS/1% TETRONIC 908
PBS/BSA (Control)	$3 \times 10^{-4}$	$1 \times 10^{-4}$
PBS/PVA /BRIJ 30 (Invention)	$7 \times 10^{-5}$	$3 \times 10^{-5}$

### EXAMPLE 12

#### Effect of Various Surfactants Added to the Solid-Phase Reagent on Non-Specific Binding of BgG to Polystyrene-Coated Magnetic Particles

The protocol for this study has been described in Example 11. The wash solution for all samples of this study was PBS/PVA/BRIJ 30. Results are summarized in TABLE 9.

TABLE 9

Fractional Non-Specific Binding of AE-Labeled BgG to Magnetic Polystyrene Microspheres		
Surfactant in PBS/PVA Wash Reagent	Surfactant/Polymer in PBS Buffered Solid-phase Reagent	Fractional Non-Specific Binding
0.1 wt.% BRIJ 30	None	$9.6 \times 10^{-5}$
0.1 wt.% BRIJ 30	1 w/v% TWEEN 20	$1.9 \times 10^{-4}$
0.1 wt.% BRIJ 30	1 w/v% TRITON X-100	$3.3 \times 10^{-4}$
0.1 wt.% BRIJ 30	1 w/v% PLURONIC F108	$4.0 \times 10^{-5}$
0.1 wt.% BRIJ 30	1 w/v% TETRONIC 908	$3.5 \times 10^{-5}$
0.1 wt.% BRIJ 30	1 w/v% PEO	$8.8 \times 10^{-6}$
0.1 wt.% BRIJ 30	1 w/v% PVA	$4.7 \times 10^{-5}$

### Claims

1. An assay for detecting or measuring an analyte in a sample characterised in that it comprises:

(1) in a heterogeneous specific binding assay, contacting the sample with (a) a tracer reagent comprising a labeled analyte analogue or a labeled analyte; and (b) solid-phase reagent comprising a solid-phase having attached thereto a specific binding partner to the said analyte or analyte analogue, to form an immobilized specific binding reaction product complex comprising the said labeled analyte analogue or the said labeled analyte paired with the said immobilized specific binding partner;

or

in a heterogeneous specific binding assay, contacting the sample with (a) a tracer reagent comprising a labeled specific binding partner to the analyte; and (b) a solid-phase reagent comprising a solid-phase having attached thereto an analyte or an analyte analogue, to form an immobilized specific binding reaction product complex comprising the said labeled specific binding partner paired with the said immobilized analyte or analyte analogue;

or

in a specific binding sandwich assay, contacting the sample with (a) a tracer reagent comprising a labeled specific binding partner to the analyte; and (b) a solid-phase reagent comprising a solid-phase having attached thereto an unlabeled specific binding partner to the analyte to form an immobilized specific binding reaction product complex comprising the said labeled specific binding partner paired with the said analyte paired with the said immobilized unlabeled specific binding partner;

(2) separating the said solid-phase having the immobilized complex thereon from the said tracer reagent by a method comprising employing a wash reagent;

(3) directly or indirectly detecting the said label on the said immobilized complex; and

(4) correlating the said detected label with the presence of the said analyte in the said sample;

a surfactant selected from a polyoxyethylene-alkylether, a polyalkylene oxide-modified polydimethylsiloxane block copolymer, a polyalkylene oxide-modified polymethylsiloxane block copolymer or a mixture thereof being present in one or more reagents selected from the said wash reagent, tracer reagent or solid-phase reagent in an effective amount to reduce non-specific binding in the said assay.

2. An assay as claimed in claim 1 wherein the said surfactant is used to pretreat the solid-phase prior to the attachment of the specific binding partner thereto, or the said surfactant is included in the solid-phase reagent after the specific binding partner has been attached thereto, or the said surfactant is used to pretreat the solid-phase prior to the attachment of the specific binding partner thereto and is included in the solid-phase reagent after the specific binding partner has been attached thereto;

or the said surfactant is present in the said tracer reagent;

or the said surfactant is present in the said wash reagent;  
or the said surfactant is present in the said wash reagent, the said tracer reagent and the said solid-phase reagent.

- 5 3. An assay as claimed in claim 1 or claim 2 wherein the said surfactant is present in an amount of from 0.001 to 3 w/v %, preferably from 0.005 to 1.5 w/v %, for each reagent.
- 10 4. An assay as claimed in any of claims 1 to 3 wherein the said surfactant present in the said wash reagent is selected from a polyoxyethylene lauryl ether (preferred) and a polyalkylene oxide modified polymethylsiloxane, and the said surfactant present in the said tracer reagent is a polyalkylene oxide modified polydimethylsiloxane.
- 15 5. An assay as claimed in claim 4 wherein the said surfactant(s) is/are employed in an amount of from 0.01 to 1 w/v % for each reagent and the said tracer reagent further comprises phosphate buffer and bovine serum albumin.
6. An assay as claimed in any of claims 1 to 5 wherein the said non-specific binding is reduced at least 1000 fold.
7. An assay as claimed in any of claims 1 to 6 wherein the said specific binding partner is an antibody or an oligonucleotide.
- 20 8. A method for reducing non-specific binding in a pseudo-homogenous assay characterised in that it comprises employing a surfactant selected from a polyoxyethylene-alkylether, a polyalkylene oxide-modified polydimethylsiloxane block copolymer, a polyalkylene oxide-modified polymethylsiloxane block copolymer or a mixture thereof in an effective amount, preferably from 0.001 to 3 w/v %, to reduce non-specific binding in the said assay.
- 25 9. A test kit for immunoassays and gene probe assays characterised in that it comprises (a) a solid-phase reagent; (b) a tracer reagent; and (c) a wash reagent; at least one of the said reagents including from 0.001 to 3 w/v % of a surfactant selected from a polyoxyethylene-alkylether, a polyalkylene oxide-modified polydimethylsiloxane block copolymer, a polyalkylene oxide-modified polymethylsiloxane block copolymer or mixtures thereof.
- 30 10. A test kit as claimed in claim 9 wherein it further comprises (d) a calibrator solution comprising a known amount of the said analyte.

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**(54) Specific binding assays and reagents therefore**

(57) A sensitive assay method has been discovered that reduces the amount of non-specific binding present in an assay, the method comprising detecting an analyte present in a sample through a specific binding reaction in which either an analogue of the analyte or a specific binding partner of the analyte is immobilized on a solid-phase and said specific binding reaction produces a detectable product immobilized on said solid-phase that

may be correlated to the amount of analyte present in the sample wherein said assay employs an effective amount of a surfactant selected from the group consisting of a polyoxyethylene-alkylether, a polyalkylene oxide-modified polydimethylsiloxane block copolymer, a polyalkylene oxide-modified polymethylsiloxane block copolymer, and mixtures thereof to reduce non-specific binding.

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# EUROPEAN SEARCH REPORT

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DOCUMENTS CONSIDERED TO BE RELEVANT			
Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim	CLASSIFICATION OF THE APPLICATION (Int.Cl.6)
X	WO-A-90 09594 (EASTMAN KODAK COMPANY) 23 August 1990 * page 15, line 5 - line 13 * ---	1-9	G01N33/543 //G01N33/573
X	EP-A-0 133 272 (BOEHRINGER MANNHEIM GMBH) 20 February 1985 * page 3, line 16 - line 28 * ---	1-9	
X	DATABASE WPI Week 9311 Derwent Publications Ltd., London, GB; AN 93-089297 XP002003294 & JP-A-05 034 353 (FUJI PHARM IND CO LTD) , 9 February 1993 * abstract * ---	1-9	
A	EP-A-0 217 246 (MILES LABORATORIES, INC.) 8 April 1987 * the whole document * -----	1	
			TECHNICAL FIELD SEARCHED (Int.Cl.6)
			G01N C12Q
The present search report has been drawn up for all claims			
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<p><b>CATEGORY OF CITED DOCUMENTS</b></p> <p>X : particularly relevant if taken alone  Y : particularly relevant if combined with another document of the same category  A : technological background  O : non-written disclosure  P : intermediate document</p> <p>T : theory or principle underlying the invention  E : earlier patent document, but published on, or after the filing date  D : document cited in the application  L : document cited for other reasons  -----  &amp; : member of the same patent family, corresponding document</p>			

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### CLAIMS INCURRING FEES

The present European patent application comprised at the time of filing more than ten claims.

- ☐ All claims fees have been paid within the prescribed time limit. The present European search report has been drawn up for all claims.
- ☐ Only part of the claims fees have been paid within the prescribed time limit. The present European search report has been drawn up for the first ten claims and for those claims for which claims fees have been paid, namely claims:
- ☐ No claims fees have been paid within the prescribed time limit. The present European search report has been drawn up for the first ten claims.

### ☒ LACK OF UNITY OF INVENTION

The Search Division considers that the present European patent application does not comply with the requirement of unity of invention and relates to several inventions or groups of inventions, namely:

1. Claims 1-10 partly: An assay and kit for detecting an analyte in a sample characterised in that it comprises a polyoxyethylene-alkylether as surfactant.
2. Claims 1-10 partly: An assay and kit for detecting an analyte in a sample characterised in that it comprises a surfactant selected from a polyalkylene oxide-modified polydimethylsiloxane block copolymer or a polyalkylene oxide-modified polymethylsiloxane block copolymer.

- ☒ All further search fees have been paid within the fixed time limit. The present European search report has been drawn up for all claims.
- ☐ Only part of the further search fees have been paid within the fixed time limit. The present European search report has been drawn up for those parts of the European patent application which relate to the inventions in respects of which search fees have been paid, namely claims:
- ☐ None of the further search fees has been paid within the fixed time limit. The present European search report has been drawn up for those parts of the European patent application which relate to the invention first mentioned in the claims, namely claims: